

International Journal of Plant & Soil Science

Volume 35, Issue 15, Page 40-47, 2023; Article no.IJPSS.99788 ISSN: 2320-7035

In vitro Evaluation of Antifungal Activity of Different Trichoderma spp. and Plant Extracts against Sclerotinia sclerotiorum (Lib.) de Bary Causing Stem Rot of Mustard

Md Zulkar Nain ^a, R. U. Khan ^a, Vaibhav Pratap Singh ^{a*} and Sibte Sayyeda ^a

^a Department of Plant Protection, Faculty of Agricultural Sciences, Aligarh Muslim University, Aligarh-202002. India.

Authors' contributions

This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.

Article Information

DOI: 10.9734/IJPSS/2023/v35i153072

Open Peer Review History:

This journal follows the Advanced Open Peer Review policy. Identity of the Reviewers, Editor(s) and additional Reviewers, peer review comments, different versions of the manuscript, comments of the editors, etc are available here:

https://www.sdiarticle5.com/review-history/99788

Original Research Article

Received: 08/03/2023 Accepted: 11/05/2023 Published: 09/06/2023

ABSTRACT

Sclerotinia stem rot (SSR) is one of the most destructive disease of mustard which affects the quality and quantity of the crop and resulting in severe yield losses. The present study was conducted to evaluate the bio-efficacy of different *Trichoderma* spp. and plant extracts against *Sclerotinia sclerotiorum* under laboratory conditions. All the tested *Trichoderma* spp. and plant extracts showed a significant effect on growth inhibition of test fungus over control. Among different *Trichoderma* spp., maximum growth inhibition (73.33%) was observed with *T. viride* followed by *T. harzianum* (67.77%). While as the least efficacy was found in *T. atroviride* (38.37%) against the pathogen. Of all, *Allium sativum* was proved to be most potent at all three tested concentrations (5,

*Corresponding author: E-mail: vaibs2121@gmail.com;

10, and 15%), thereby registering 59.26, 67.41, and 100.0% growth inhibition followed by *Azadirachta indica* and *Lantana camara*, respectively. However, *Argemone maxicana* was least effective at all concentrations in this study.

Keywords: Trichoderma spp.: plant extracts: rapeseed-mustard: sclerotinia stem rot: in vitro.

1. INTRODUCTION

India is one of the leading edible oil economy after the USA. China, and Brazil, which contributes about 10 per cent to the global oilseed oil. Rapeseed-mustard is the third major oilseed crop after soybean and palm in the world, and about 25 per cent of total oilseed produced by this crop [1,2]. This crop used as an important source of edible oil, industrial oil, livestock feed, vegetable, and soil amendment. The mustard has nutritional value, viz., sugar 1.41g, carbohydrates 4.51g, fat 0.47g, protein 2.56g per 100 g (3.5 oz), and dietary fiber 2g. India holds second place in the area (20.50%) and fourth rank in yield production (10.72%) of rapeseed and mustard, contributing 6.23 m ha and 9.34 mt, respectively, with an average productivity of 1499 kg ha⁻¹ [3]. A wide gap occurs between the probable yield and the actual yield at the farmer's field, which is mainly because of biotic and abiotic factors to which this crop is exposed. Amongst, diseases caused by fungi are posing a severe menace. These fungal diseases cause deterioration in oilseed quality also reduce production. About thirty diseases are known to occur on crops in India [4]. Some of the economically critical diseases of rapeseed-mustard reported in India or abroad are white rust, downy mildew, Alternaria blight, stem rot, powdery mildew, Rhizoctonia leaf blight, black leg/canker, grey mold, Fusarium wilt, damping off, black rot and mosaic [5].

Amongst, Sclerotinia stem rot (SSR) has become one of the most ubiquitous and devastating soilborne disease distributed worldwide. disease occurs commonly in cool and wet regions on a number of plants that predominantly grown in temperate and subtropical regions all over the world [6-8]. Earlier, this disease was considered to be of lesser importance, but due to newer crop practices and large cultivation of susceptible cultivars by the farmers, it has now achieved the next place to Alternaria blight in their economic importance [9,10]. In India, this disease has been reported in serious proportion in states like Uttar Pradesh, Rajasthan, Madhya Pradesh, Punjab, Haryana, and Bihar [11]. In Rajasthan, about 60 per cent yield loss has been observed in heavily infected plants [12]. According to Shukla [13], plants infected before initiation of flower can result in 100 per cent crop losses, whereas the plants attacked after flowering suffer 50 per cent yield loss.

The pathogen appears to be non-specific and attacks different plant parts, viz., leaf, stem, and pods. The characteristic symptoms appeared as elongated water-soaked lesions at stem base, which usually enlarge quickly, become necrotic. and subsequently develop patches of white fluffy mycelium [14]. Later, the stem is girdled completely and covered by a cottony mycelium growth; it breaks from where it shows rotting and drying. The pathogen survives through latent sclerotia which can remain viable for relatively longer time as they are resistant to adverse conditions (physically and chemically), as well as degradation another by beneficial microorganism [15].

The management of Sclerotinia rot is absolutely difficult because of the broad host range of the pathogen, variability, and soil-inhabitant nature [16,17]. The management of stem rot needs a proper understanding of pathogen etiology and epidemiology. Application of fungicides is the most popular method used by the farmers for the management of S. sclerotiorum. But extensive use of fungicides causes hazardous effect on health of human, animals and also pollutes the environment. So the best alternative fungicides is use of plant extracts and bio-control agents because it is cheap, effective, and environmentally safe. Keeping in mind the significance of crop, the current experiment was conducted in vitro to the effectiveness of different Trichoderma spp. and plant extracts in managing the disease.

2. MATERIALS AND METHODS

2.1 *In vitro* Evaluation of Antagonistic Effect of *Trichoderma* spp. against *S. sclerotiorum*

Four *Trichoderma* spp. namely, *T. harzianum*, *T. atroviride*, *T. viride*, and *T. virens* were tested to check their inhibitory effect on the growth of *S. sclerotiorum* using a dual culture method (Dennis

and Webster, 1971). Twenty ml of sterilized medium (PDA) was transferred into a sterile Petri plate and allowed for solidification. Five mm diameter discs from fresh colonies of the pathogen were cut with the help of a sterile cork borer and positioned near the periphery of the plate having PDA. Similarly, Trichoderma spp. was also placed on another side, i.e., at a 180° angle. Plates with no antagonists are treated as a control for the pathogen. Each treatment was replicated thrice and incubated at 25±2°C for seven days. The antagonistic activity by Trichoderma spp. was recorded after an incubation period of 7 days by assessing the growth of the fungus in both treated and untreated plates. The growth inhibition of the pathogen was measured by using the method given by Vincent [18].

Per cent inhibition (%) = $(C - T)/C \times 100$

Where,

C=pathogen growth in control plate T=pathogen growth in dual culture plate

2.2 In vitro Evaluation of Antifungal Activity of Plant Extracts against S. sclerotiorum

Antifungal activity of four plant extracts i.e., Neem (Azadirachta indica), Lantana (Lantana camara), Satyanashi (Argemone maxicana) and Garlic (Allium sativum) were also evaluated against S. sclerotiorum at three different concentrations (5%, 10% and 15%) by poison food technique [19]. About 100 gm of fresh leaves/clove were used and carefully washed in distilled water. Such plant parts were cut into small pieces and then grinded in a mortar and pestle by including 100 ml of distilled water. The basic material was then filtered through muslin cloth (double layer), and then the filtrate was filtered through Whatman no. 1 filter paper. The prepared plant extracts were autoclaved at temperature of 40°C for 5 minutes to prevent contamination.

The requisite amount of plant extracts was incorporated into sterilized potato dextrose agar medium (non-solidified) and stirred well to make it standardized. Thereafter, 20 ml of amended medium was then poured into 90 mm measuring Petri plates. The observation, thus, recorded the growth of fungus at all tested concentrations until the growth of test fungus fully covered the unpoisoned Petri plates (check). The percent

inhibition in radial growth (T) over control (check) was assessed by using the following method given by Vincent, [18].

Per cent inhibition (%) = $(C - T)/C \times 100$

Where.

C= growth of the pathogen in control plates T= growth of the pathogen in treated plates

3. RESULTS AND DISCUSSION

3.1 *In-vitro* Efficacy of *Trichoderma* spp. against *S. sclerotiorum*

All *Trichoderma* spp. showed a significant impact on radial growth inhibition of test fungus over control. Among all tested *Trichoderma* spp., significant maximum growth inhibition (73.33%) was recorded due to *T. viride* which was followed by *T. harzianum* (67.77%), *T. virens* (44.44%). While as, the least efficacy was found in *T. atroviride* (38.37%) against the pathogen (Fig. 1, Plate 1).

It is clear from the present observations that among all Trichoderma spp., T. viride was the most efficient in radial growth inhibition of the fungus followed by T. harzianum, T. virens, while T. atroviride was the least effective in this study (Fig. 1, Plate 1). The present findings are in testimony to the results of several earlier reports that also indicated the efficacy of T. virens, T. viride, and T. harzianum against S. sclerotiorum in reducing the mycelial growth and sclerotial development [20-22]. In a study, Mehta et al. [23] also tested the effect of several antagonists against S. sclerotiorum (stem rot of mustard) and reported that *T. harzianum* was most superior as compared to other strains of Trichoderma in vitro and it was also found to be excellent in checking length of lesions and post emergence dampingoff in vivo. The effectiveness of bio-control agents in minimizing the mycelial growth and sclerotial development against S. sclerotiorum has also been reported by several scientists and researchers [23,24].

Different *Trichoderma* spp. are known to suppress *S. sclerotiorum* by coiling their hyphae rather than sclerotia by mechanisms involved mycoparasitism, antibiosis, and systemically induced resistance [25,26]. For parasitizing the pathogen, a number of enzymes like glucanases, chitinases, cellulases and proteases are produced by *Trichoderma* spp. which results in disintegration of fungal cell wall [27].

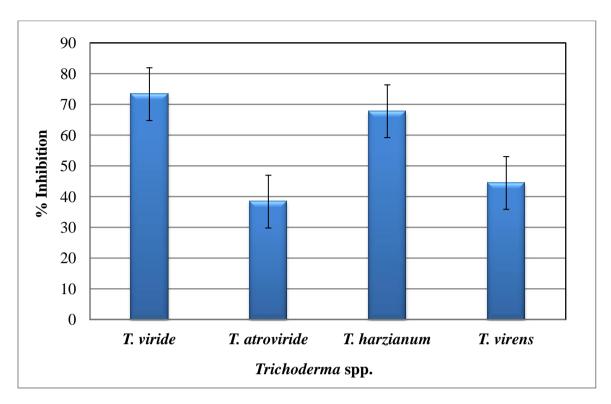


Fig. 1. Efficacy of different Trichoderma spp. on growth inhibition of S. sclerotiorum

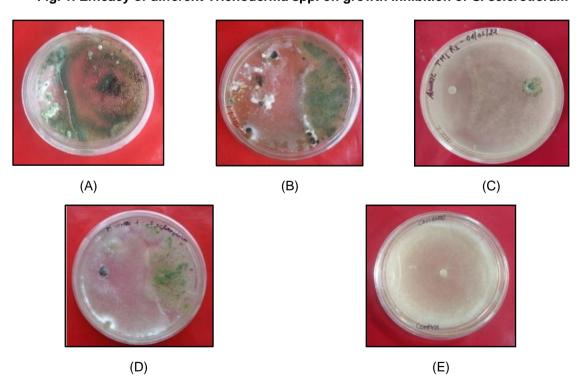


Plate 1. Efficacy of different *Trichoderma spp.* on growth of S. sclerotiorum, T. viride (A), T. atroviride (B), T. harzianum (C), T. virens (D) and control (E)

3.2 *In vitro* Efficacy of Different Plant Extracts against *S. sclerotiorum*

All plant extracts significantly reduced the growth of fungus at all concentrations over control. Of all, *Allium sativum* was found to be most effective at all three concentrations of 5, 10, and 15 per cent, thereby registering 59.26, 67.41, and 100 per cent radial growth inhibition,

followed by Azadirachta indica (51.11, 65.55, and 82.6%) and Lantana camara (43.33, 64.44 and 77.77%), respectively. However, Argemone maxicana was least effective at all concentrations in comparison to other plant extracts and resulted in 33.33, 51.48, and 63.33 per cent inhibition of growth at 5, 10, and 15 per cent concentrations, respectively (Table 1, Plate 2).

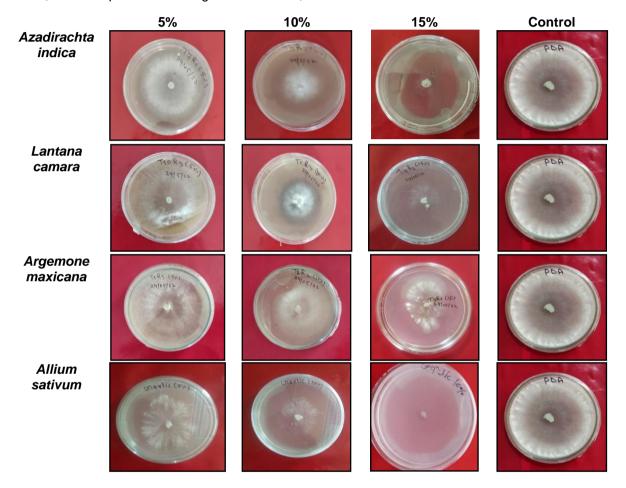


Plate 2. Effect of different plant extracts on growth of S. sclerotiorum

Table 1. Effect of different plant extracts on growth inhibition of S. sclerotiorum

Plant extracts	Concentrations					
	5%		10%		15%	
	Radial growth	%	Radial	%	Radial	%
	(mm)	Inhibition	growth (m	nm) Inhibition	growth	(mm) Inhibition
Azadirachta indica	51.00	51.11 ^b	31.00	65.55 ^d	15.66	82.6 ^b
Lantana camara	36.66	43.33 ^c	32.00	64.44 ^c	20.00	77.77 ^c
Argemone maxicana	60.00	33.33 ^d	43.66	51.48 ^b	33.00	63.33 ^d
Allium sativum	44.00	59.26 ^a	29.33	67.41 ^a	00.00	100.0 ^a
Control	90.00	-	90.00	-	90.00	-
L.S.D. (P≤0.05)	4.28	4.76	3.54	3.93	2.83	3.14
S.E(m)±	0.94	1.04	0.78	0.86	0.62	0.69

^{*}Each value is an average of three replicates. Values within a column followed by different alphabets are significant and some alphabets are non-significant according to Tukey's Test at P≤0.05

It is, thus, clear from this study Allium sativum was the most effective plant extracts, followed by *indica* and Azadirachta Lantana However, Argemone maxicana was the least effective in this study (Table 1, Plate 2). The results, thus, obtained in the present study are in agreement with the findings of other researchers, who have also noted the efficacy of garlic and neem extracts for inhibition of mycelial growth and sclerotial formation of S. sclerotiorum [28-32.21]. In a study, Fagodia et al. [33] also investigated the efficacy of different botanicals on the growth of S. sclerotiorum causing stem rot of coriander and found that Allium sativum extract was most potent in inhibiting mycelial growth of fungus, followed by Eucalyptus leaf extract [34].

4. CONCLUSION

All *Trichoderma* spp. significantly inhibited the growth of *S. sclerotiorum*. However, *T. viride* and *T. harzianum* was noted to be most effective antagonist in inhibiting the maximum radial growth of the pathogen. While as, *T. atroviride* proved to be least efficacious in this study. Of all plant extracts, *Allium sativum* was found superior over all others at all concentrations followed by *Azadirachta indica* and *Lantana camara*. However, *Argemone maxicana* was found least effective in this study. The biopesticides should be used by the farmers alone or in combinations with other cultural practices because it is economical and ecofriendly than chemicals.

CONFERENCE DISCLAIMER

Some part of this manuscript was previously presented in the conference: 3rd International Conference IAAHAS-2023 "Innovative Approaches in Agriculture, Horticulture & Allied Sciences" on March 29-31, 2023 in SGT University, Gurugram, India. Web Link of the proceeding: https://wikifarmer.com/event/iaahas-2023-innovative-approaches-in-agriculture-horticulture-allied-sciences/

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Yadav MS, Godika S, Yadava DK, Ahmad N, Mehta N, Bhatnagar K, Agrawal VK,

- Kumar A, Thomas L, Chattopadhyay C. Prioritizing components of package of integrated pest management in Indian mustard (*Brassica juncea*) in India for better economic benefit. Crop Protection. 2019;120:21-29.
- Chand S, Prakash Patidar O, Chaudhary R, Saroj R, Chandra K, Kamal Meena V, Vasisth P. Rapeseed-mustard breeding in India: Scenario, achievements and research needs. Intech Open; 2021. DOI: 10.5772/intechopen.96319
- Agricultural Statistics at a Glance. Directorate of Economics and Statistics Department of Agriculture and Farmers Welfare Ministry of Agriculture and Farmers Welfare Government of India; 2021.
- Saharan GS. Management of rapeseed and mustard diseases. In: Advances in oilseed research. Rapeseed and Mustard (Kumar D. and Rai M, eds.) Scientific Pub. Jodhpur. 1992;1:155-188.
- Saha LR, Singh HB, Raychaudhuri SP, Verma JP. Diseases of rapeseed and mustard and their management. Rev. Tropical Pl. Pathol. 1989;5:47-77.
- 6. Purdy LH. *Sclerotinia sclerotiorum*: History, diseases and symptomatology, host range, geographic distribution, and impact. Phytopathology. 1979;69:875-880.
- 7. Saharan GS, Mehta N. Sclerotinia diseases of crop plants: Biology, ecology and disease management: Springer Press. Sclerotinia sclerotiorum Isolates in Northern Parts of Iran. World Appl. Sci. J, 2008;8(3):326-333.
- Sharma P, Meena PD, Singh D. Effect of Sclerotinia sclerotiorum culture filtrate on seed germination and seedling vigour of Indian mustard (Brassica juncea cv. Rohini). J Oilseed Brassica. 2014;5:158-161.
- Kolte SJ. Diseases of annual edible oilseed crops, rapeseed-mustard and sesame diseases. CRC Press Inc Boca Raton. 1985;2:135.
- Parveen K, Haseeb A, Shukla PK. Management of Sclerotinia sclerotiorum on Mentha arvensis C.V.Gomti. J. Mycol. Pl. Pathol. 2007;37(1):33-36.
- Saharan GS, Mehta N. Fungal diseases of rapeseed-mustard. In: Diseases of Field Crops (Gupta, VK. and Paul YS. eds.). Indus Publishing Company, New Delhi. 2002:193-228.

- Ghasolia RP, Shivpuri A, Bhargava AK. Sclerotinia rot of Indian mustard (*Brassica juncea*) in Rajasthan. Indian Phytopathol. 2004;57:76-79.
- 13. Shukla AK. Estimation of yield losses to Indian mustard (*Brassica juncea*) due to sclerotinia stem rot. J Phytol Res. 2005;18:267-268.
- Bolton DM, Thomma PHJB, Nelson DB. Sclerotinia sclerotiorum (Lib.) de Bary: Biology and molecular traits of a cosmopolitan pathogen. Mol Plant Pathol. 2006;7:1-16.
- 15. Wu BM, Subbarao KV. Effects of soil temperature, moisture and burial depths on carpogenic germination of *Sclerotinia sclerotiorum* and *S. minor*. Phytopathology. 2008;98:1144–1152.
- Mondal B, Khatua DC, Hansda S, Sharma R. Addition to the host range of *Sclerotinia* sclerotiorum in West Bengal. Scholars Academic Journal of Biosciences. 2015;3:361-364.
- 17. Smolinska U, Kowalska B. Biological control of the soil-borne fungal pathogen *Sclerotinia sclerotiorum* a review. Journal of Plant Pathology. 2018;100:1-12.
- 18. Vincent JM. Distortion of fungal hyphae in the presence of certain inhibitors. Nature. 1947;159:239-241.
- 19. Dubey SC, Patel B. Evaluation of fungal antagonist against *Thanatephorus cucumeris* causing web blight urd and mung bean. Indian Phytopath. 2001;54:206–209.
- 20. Pandey P, Kumar R, Mishra P. Integrated approach for the management of *Sclerotinia sclerotiorum* (Lib.) de Bary, causing stem rot of chickpea. Indian Phytopath. 2011;64(1):37-40.
- 21. Yadav M, Rakholiya KB, Pawar DM. Evaluation of bioagents for management of the onion purple blotch and bulb yield loss assessment under field conditions. The Bioscan. 2011;8(4):1295-1298.
- 22. Sharma P, Meena PD, Singh S, Rai PK. Efficacy of micro-nutrients, fungicides and bio-agents against sclerotinia stem rot (*Sclerotinia sclerotiorum*) of Indian mustard. Int. J. Curr. Microbiol. App. Sci. 2017;6(10):620-626.
- 23. Mehta N, Hieu N, Sangwan M. Efficacy of various antagonistic isolates and species of against causing white stem rot of mustard. Journal of Mycology and Plant Pathology. 2012;42(2):244-250.

- 24. Shivpuri A, Gupta RBL. Evaluation of different fungicides and plant extracts against *Sclerotinia sclerotiorum* causing stem rot of mustard. Phytopathology. 2001;52:272-274.
- 25. Vinale F, Sivasithamparam K, Ghisalberti EL, Marra R, Woo SL, Lorito M. *Trichoderma*–plant–pathogen interactions. Soil Biology and Biochemistry. 2008;40(1):1-10.
- Geraldine A, Fabyano L, Carvalho C, Diego D, Elder B, Amanda R, Ulhoa C, Murillo LJ. Cell wall-degrading enzymes and parasitism of sclerotia are key factors on field biocontrol of white mold by *Trichoderma* spp. Biological Control. 2013;67:308-316.
- López-Mondéjar R, Ros M, Pascual J. Added-value of *Trichoderma* amended compost as biopesticide organic substrates: Alternative to traditional organic substrates. Acta Horticulturae. 2011;898:189-196.
- 28. Kapil R, Kapoor AS. Management of white rot of pea incited by *Sclerotinia sclerotiorum* using *Trichoderma* spp. and biopesticides. Indian Phytopathology. 2005;58:10-16.
- 29. Tripathi AK, Tripathi SC. Management of sclerotinia stem rot of Indian mustard through plant extracts. Vegetos. 2009;22(1):1-3.
- Mehta N, Hieu NT, Sangwan MS. Efficacy of some botanicals against Sclerotinia sclerotiorum inciting white stem rot of rapeseed-mustard. Plant Dis Res. 2011;26:82-86.
- 31. Meena PD, Gour RB, Gupta JC, Singh HK, Awasthi RP, Netam RS, Godika S, Sandhu PS, Prasad R, Rathi AS, Rai D, Thomas L, Patel GA, Chattopadhyay C. Non-chemical agents provide tenable, eco-friendly alternatives for the management of the major diseases devastating Indian mustard (*Brassica juncea*) in India. Crop Protection. 2013; 53:169-174.
- 32. Fatehpuria PK, Sasode RS, Pandya RK, Singh R, Gupta JC. Efficacy of different inoculation techniques for testing the pathogenicity of *Sclerotinia sclerotiorum* causing sclerotinia blight of *Brassica juncea*. IJCS. 2017;5(5):1937-1940.
- Fagodia RK, Godika S, Fagodia BL. Effect of physical parameters on the growth and sclerotia formation of Sclerotinia

sclerotiorum (Lib.) de Bary, causing stem rot of coriander. Annals of Plant and Soil Research. 2017;19(1):54–58.

34. Dennis C, Webster J. Antagonistic

properties of species groups of *Trichoderma* - III. Production of volatile antibiotics. Trans. Br. Mycol. Soc. 1971;51:363-369.

© 2023 Nain et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history:
The peer review history for this paper can be accessed here:
https://www.sdiarticle5.com/review-history/99788